



The Effects of Bisphenol A (BPA) on the Neural Development of the *Xenopus laevis* Tadpole

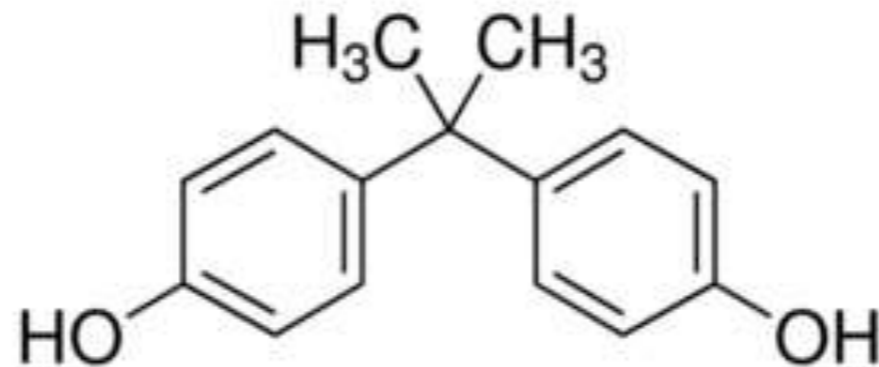
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Overview

- Introduction & Background
- Visually Guided Behavior Study
- Electrophysiology Studies
- Axon Circuit Development Study
- Conclusion
- References

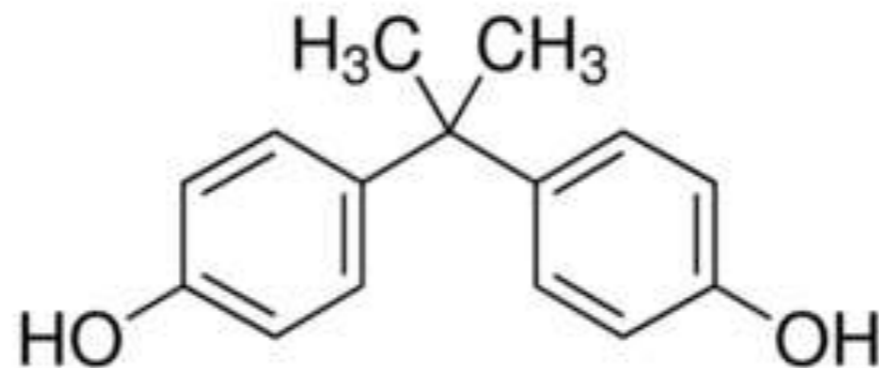
Introduction

- ▶ Bisphenol-A
 - Common chemical compound used in manufacture of plastics and canned goods
 - Known endocrine disruptor
 - 2 phenol rings - estrogen mimic



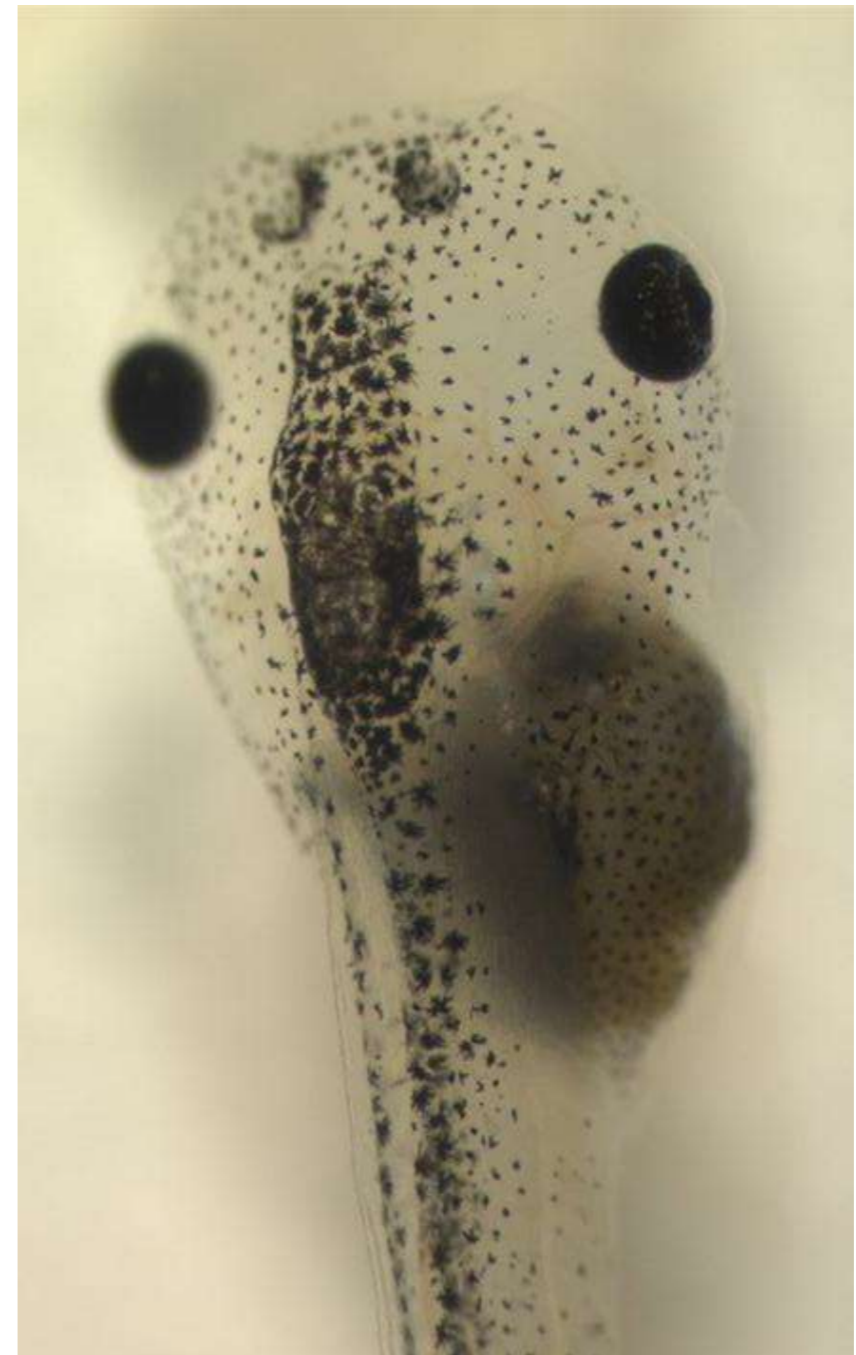
Introduction

- ▶ Bisphenol-A
 - Many studies show BPA has effects on adult mammalian nervous system
 - We are investigating BPA's effects on a developing neural circuit

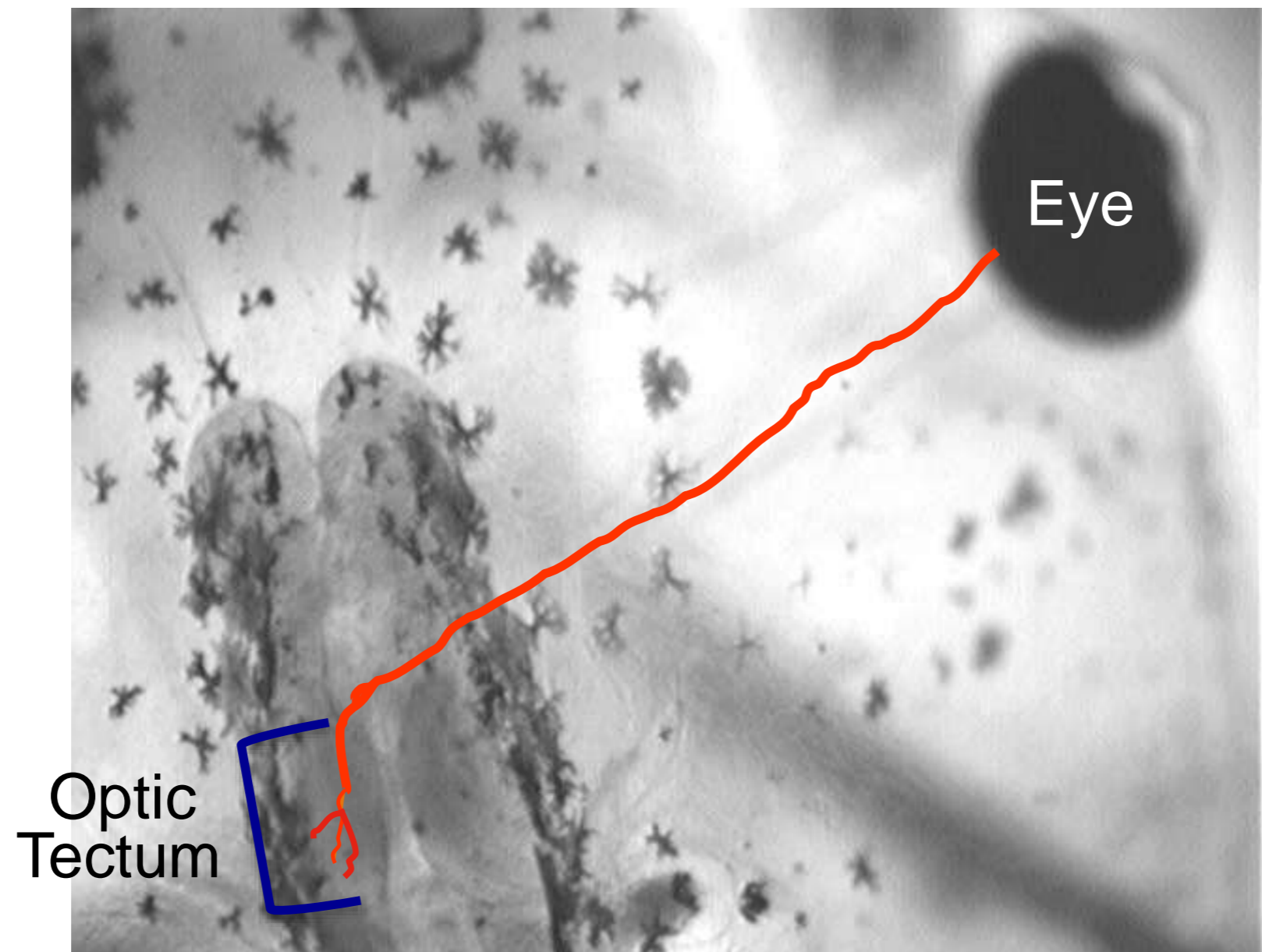


Introduction

- ▶ *Why Xenopus?*
 - Mature in relatively short time period
 - Many tadpoles per fertilization
 - Well-documented stages of development
 - Simple visual circuit model
 - Relative ease in toxin exposure and dose control



The Retinotectal is the major circuit of the tadpole visual system



Introduction

Pratt Lab - Goals

- Expose *Xenopus* to 10 & 15 μM BPA in Steinberg solution
- Divided into three parts:
 - ★ Behavior study
 - ★ Electrophysiology study
 - ★ Imaging study

Research Design

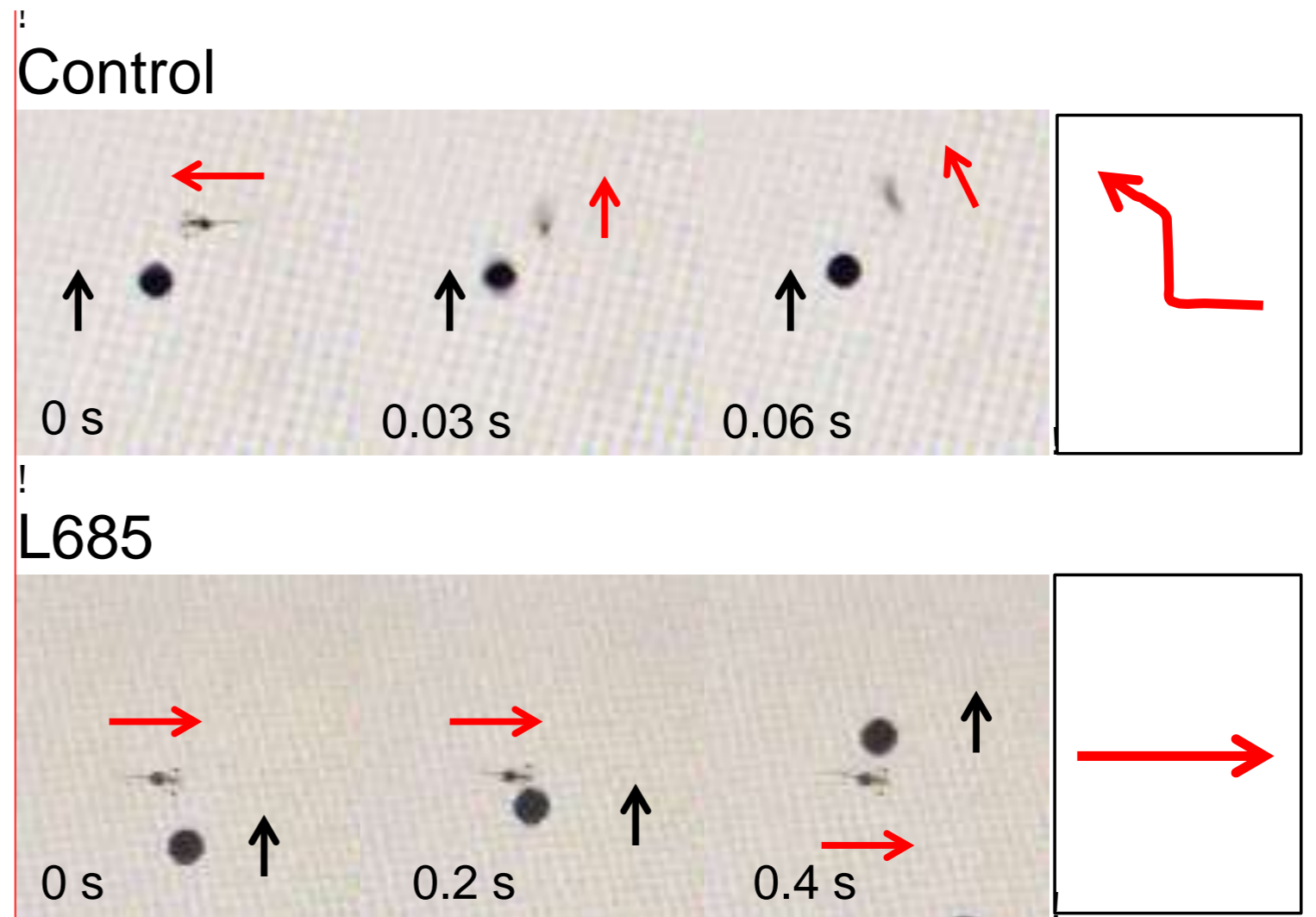
- Groups of 10 tadpole embryos were selected 12 to 24 hours post-fertilization.
- Each group was assigned to either a control, 10 μ M, or 15 μ M bowl and placed in their respective concentrations.
- The concentrations were optimized at 10 & 15 μ M.
- 100mL of Steinberg Solution in glass bowls was the synthesized habitat for the tadpoles.

Research Design

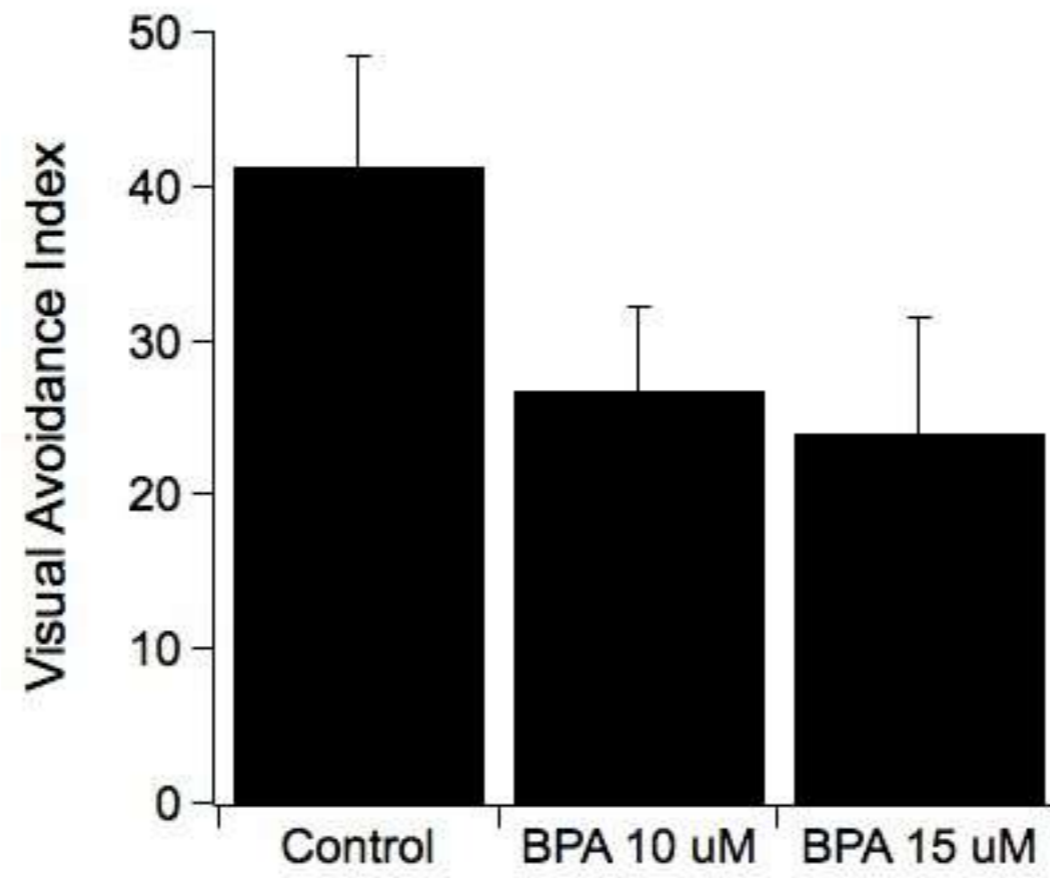
- For the first half of the lab's experiments, the tadpoles were observed every third day.
- The control and BPA solutions were refreshed every third day, using the same BPA stock for the entirety of those groups' respective experiments.

Behavioral Test Design

- Behavioral tests were conducted 8-10 days post fertilization.
- The tadpoles were recorded for three minutes, the video was analyzed for encounters the tads had with the dots.
- Five encounters for a tadpole are necessary in order for the information to be used.

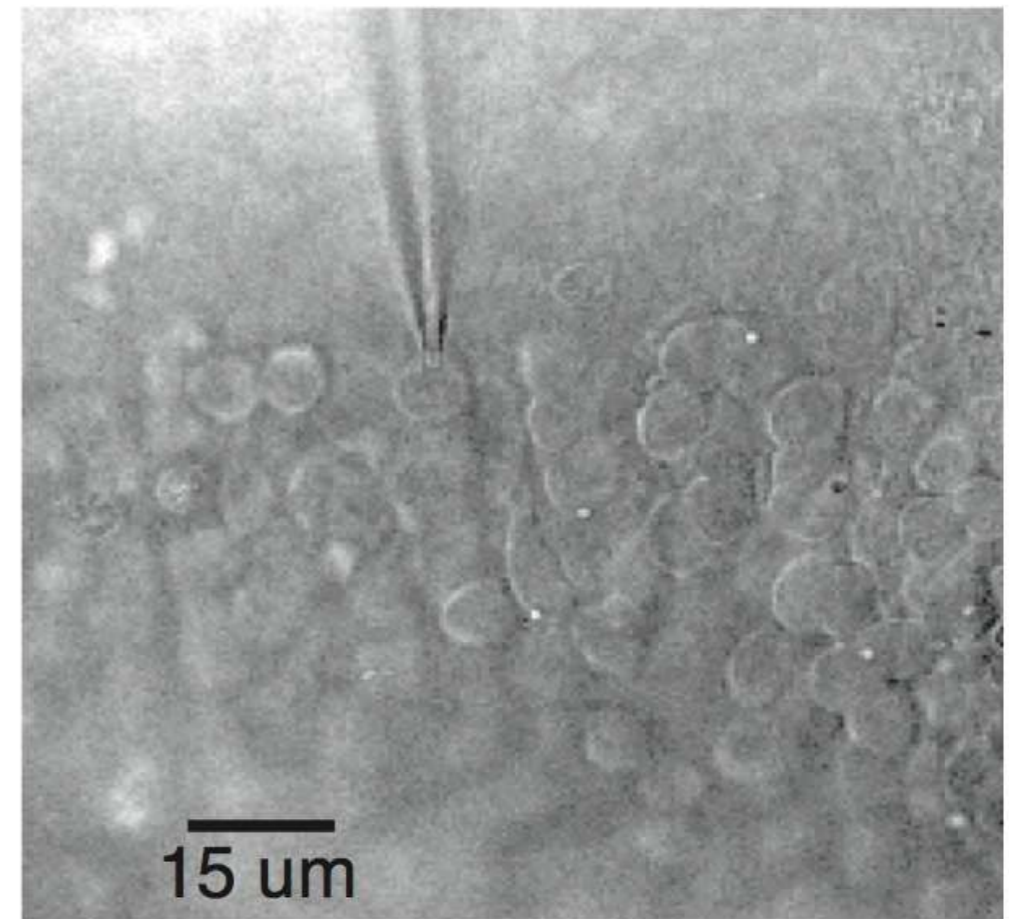


Results

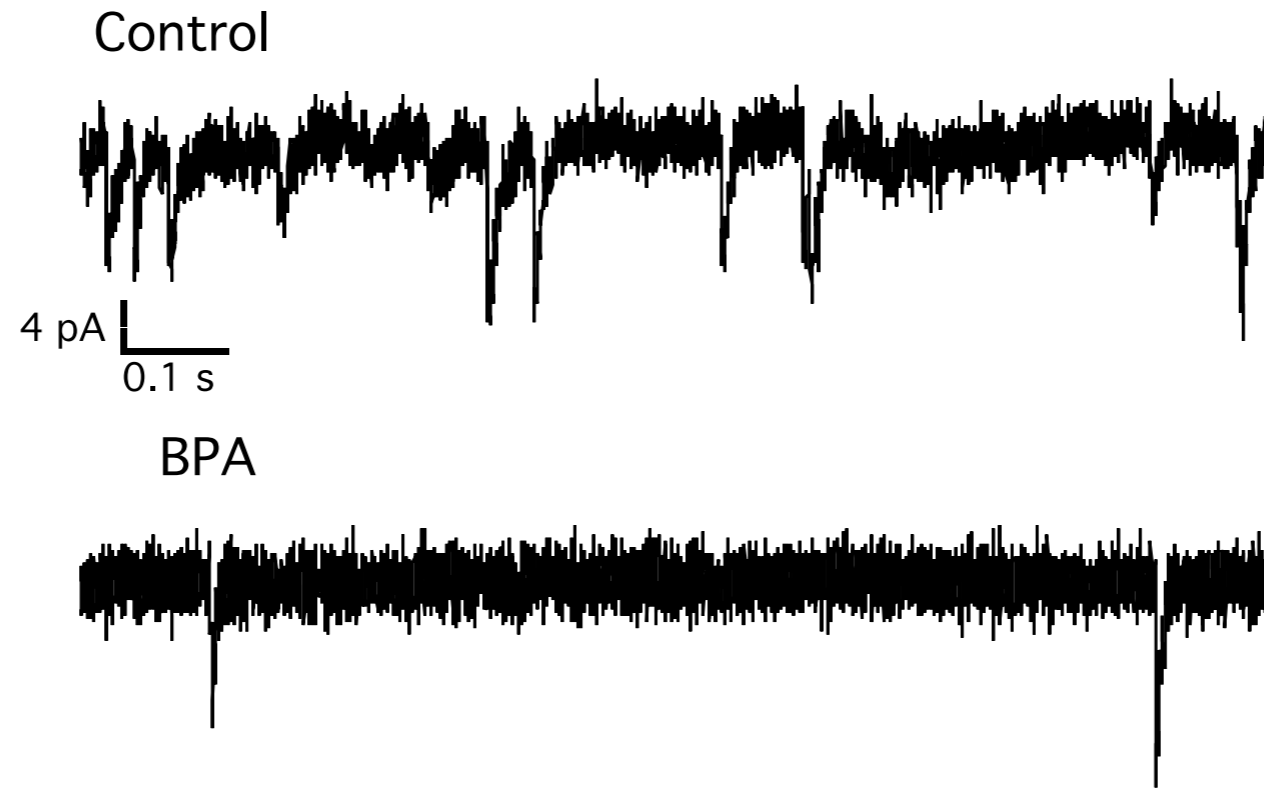


Electrophysiology Test Design

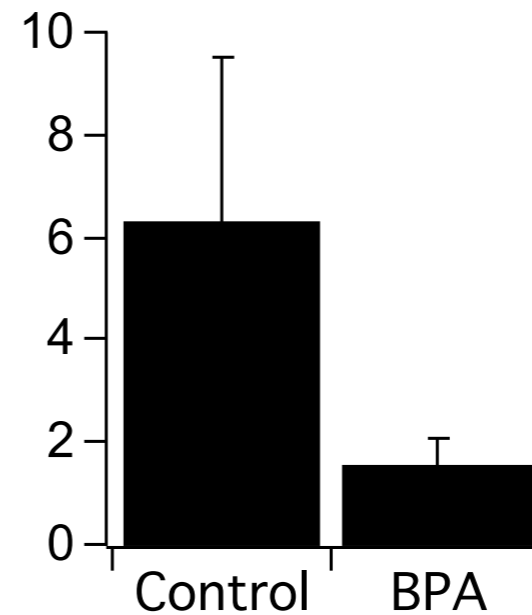
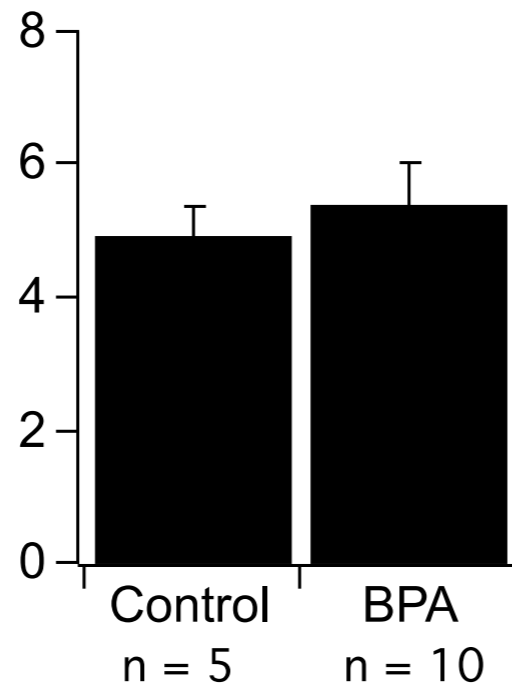
- Tadpoles at stage 48/49 were selected, 10 days post fertilization, from the 15uM concentration.
- Whole brain preparation
- Record from tectal neurons using a glass micropipette
- Recorded spontaneous Excitatory Postsynaptic Currents
 - Spontaneous activity in the brain.
 - Amplitude=Strength of Synapses
 - Frequency=Number of Synapses



Electrophysiology Results



These are 1 second traces of recordings of the spontaneous Excitatory Postsynaptic Currents (sEPSCs).



Using Fluorescent Dye to Image Visual Circuit Axons

- Injections are done using stage 44 tadpoles
- Tadpole set up on dissecting scope stage after MS-222 Immersion
- Glass pipette filled with Dye-I and injected into eyeball of tadpole
- Melanocytes on top of brain are removed by scraping to allow imaging of axons entering tectum
- Tadpole placed in formaldehyde fix for one week and then imaged using confocal microscopy

Zeiss 710 Confocal Microscope



- Provided by University of Wyoming Jenkins Microscopy Facility

Results

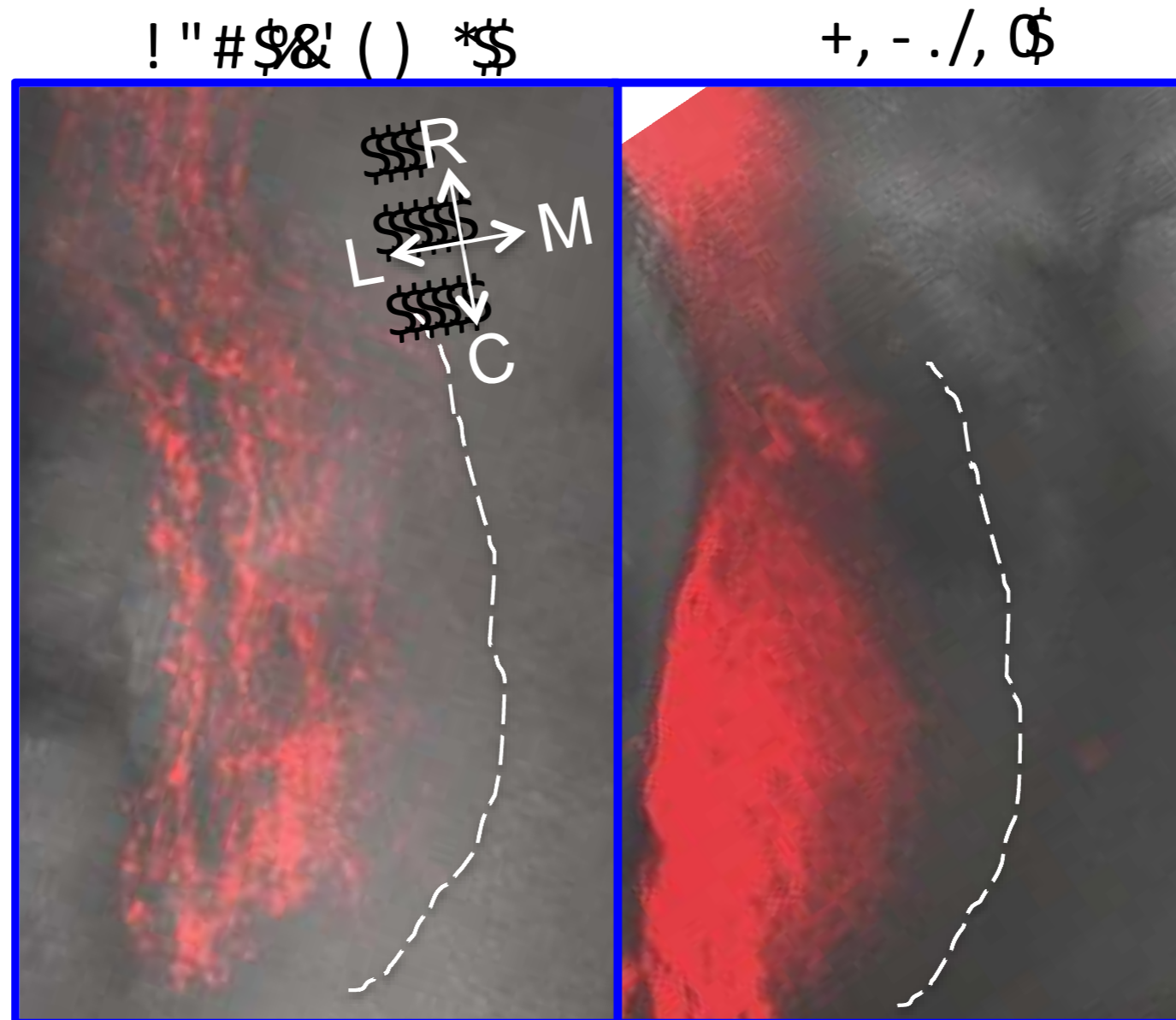


Fig. 1 - Control and BPA exposed *Xenopus laevis* tadpole retinal ganglia axons traveling to optic tectum of brain.

Future Research

- There are significant differences between BPA tadpoles and control tadpoles at the behavior, cell, and circuit levels.
- The Pratt Lab has optimized testing the visual system of tadpoles, therefore, tests at the circuit, cell, and behavioral levels will continue.
 - Every two weeks, new groups of tadpoles are tested.

Thank You

- We would like to thank INBRE and EPSCoR
- The Pratt Lab
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References

- Baba, K., Okada, K., Kinoshita, T., & Imaoka, S. (2009). Bisphenol A Disrupts Notch Signaling by Inhibiting Gamma-Secretase Activity and Causes Eye Dysplasia of *Xenopus laevis*. *Toxicological Sciences*, 108(2), 344-355.
- Deeg, K. E., Sears, I. B., & Aizenman, C. D. (2009). Development of Multisensory Convergence in the *Xenopus* Optic Tectum. *Journal of Neurophysiology*, 102(6), 3392-3404.
- Hiramoto, M., & Cline, H. T. (2009). Convergence of multisensory inputs in *Xenopus* tadpole tectum. *Developmental Neurobiology*, 69(14), 959-971.
- Imaoka, S., Mori, T., & Kinoshita, T. (2007). Bisphenol A Causes Malformation of the Head Region in Embryos of *Xenopus laevis* and Decreases the Expression of the ESR-1 Gene Mediated by Notch Signaling. *Biol. Pharm. Bull. Biological & Pharmaceutical Bulletin*, 30(2), 371-374.
- Oka, T., Adati, N., Shinkai, T., Sakuma, K., Nishimura, T., & Kurose, K. (2003). Bisphenol A induces apoptosis in central neural cells during early development of *Xenopus laevis*. *Biochemical and Biophysical Research Communications*, 312(4), 877-882.
- Paganelli, A. R., Ocaña, O. H., Prat, M. I., Franco, P. G., López, S. L., Morelli, L., . . . Carrasco, A. E. (2001). The Alzheimer-related gene presenilin-1 facilitates sonic hedgehog expression in *Xenopus* primary neurogenesis. *Mechanisms of Development*, 107(1-2), 119-131.

- Pratt, K. G., & Khakhalin, A. S. (2013). Modeling human neurodevelopmental disorders in the *Xenopus* tadpole: From mechanisms to therapeutic targets. *Disease Models & Mechanisms*, 6(5), 1057-1065.
- Ruthazer, E. S., Schohl, A., Schwartz, N., Tavakoli, A., Tremblay, M., & Cline, H. T. (2013). Dye Labeling Retinal Ganglion Cell Axons in Live *Xenopus* Tadpoles. *Cold Spring Harbor Protocols*, 2013(8), 768-770.
- Sone, K., Hinago, M., Kitayama, A., Morokuma, J., Ueno, N., Watanabe, H., & Iguchi, T. (2004). Effects of 17 β -estradiol, nonylphenol, and bisphenol-A on developing *Xenopus laevis* embryos. *General and Comparative Endocrinology*, 138(3), 228-236.
- Wu, G. (1998). Stabilization of Dendritic Arbor Structure in Vivo by CaMKII. *Science*, 279(5348), 222-226.